

# UNCLASSIFIED

AD NUMBER
ADB177265
NEW LIMITATION CHANGE
TO Approved for public release, distribution unlimited
FROM Distribution authorized to U.S. Gov't. agencies only; Premature Dissemination; 31 JUL 1993. Other requests shall be referred to U.S. Army Medical Research and Development Command, Attn: SGRD-RMI-S, Fort Detrick, Frederick, MD 21702-5012.
AUTHORITY
MCMR-RMI-S, 70-1y ltr dtd 6 Aug 1996

THIS PAGE IS UNCLASSIFIED

**AD-B177 265**



AD \_\_\_\_\_

L  
②

**CONTRACT NO: DAMD17-91-C-1006**

**TITLE: SIMIAN HEMORRHAGIC FEVER (SHF) VIRUS**

**PRINCIPAL INVESTIGATOR: Margo A. Brinton, Ph.D.**

**CONTRACTING ORGANIZATION: Georgia State University  
University Plaza  
Room 131, Sparks Hall  
Atlanta, Georgia 30303-3083**

**REPORT DATE: July 31, 1993**

**TYPE OF REPORT: Phase III Final Report**

**SDTIC  
ELECTE  
NOV 16 1993  
A**

**PREPARED FOR: U.S. Army Medical Research and  
Development Command, Fort Detrick  
Frederick, Maryland 21702-5012**

**DISTRIBUTION STATEMENT: Distribution authorized to U.S.  
Government agencies only, Premature Dissemination, July 31, 1993.  
Other requests for this document shall be referred to Commander,  
U.S. Army Medical Research and Development Command, SGRD-RMI-S,  
Fort Detrick, Frederick, Maryland 21702-5012.**

**The findings in this report are not to be construed as an  
official Department of the Army position unless so designated by  
other authorized documents.**

**93-27799**



# REPORT DOCUMENTATION PAGE

Form Approved  
OMB No. 0704-0188

Public reporting burden for this collection of information is estimated to average 1 hour per response, including the time for reviewing instructions, searching existing data sources, gathering and maintaining the data needed, and completing and reviewing the collection of information. Send comments regarding this burden estimate or any other aspect of this collection of information, including suggestions for reducing this burden, to Washington Headquarters Services, Directorate for Information Operations and Reports, 1215 Jefferson Davis Highway, Suite 1204, Arlington, VA 22202-4302, and to the Office of Management and Budget, Paperwork Reduction Project (0704-0188), Washington, DC 20503.

1. AGENCY USE ONLY (Leave blank)		2. REPORT DATE 31 July 1993	3. REPORT TYPE AND DATES COVERED Phase III Final Report	
4. TITLE AND SUBTITLE Simian Hemorrhagic Fever (SHF) Virus			5. FUNDING NUMBERS DAMD17-91-C-1006	
6. AUTHOR(S) Margo A. Brinton, Ph.D.				
7. PERFORMING ORGANIZATION NAME(S) AND ADDRESS(ES) Georgia State University University Plaza Room 131, Sparks Hall Atlanta, Georgia 30303-3083			8. PERFORMING ORGANIZATION REPORT NUMBER	
9. SPONSORING / MONITORING AGENCY NAME(S) AND ADDRESS(ES) U.S. Army Medical Research & Development Command Fort Detrick Frederick, Maryland 21702-5012			10. SPONSORING / MONITORING AGENCY REPORT NUMBER	
11. SUPPLEMENTARY NOTES				
12a. DISTRIBUTION / AVAILABILITY STATEMENT Distribution authorized to U.S. Government agencies only, Premature Dissemination, July 31, 1993. Other requests for this document shall be referred to Commander, USAMRDC, Fort Detrick, Frederick, MD 21702-5012.			12b. DISTRIBUTION CODE	
13. ABSTRACT (Maximum 200 words)  Phase III of this contract required that we twice-clone selected simian hemorrhagic fever (SHF) virus-specific hybridoma cultures, expand two clones from each clone as well as 50 ml of supernatant fluid from cultures of each clone.				
14. SUBJECT TERMS RA I, Hemorrhagic Fevers, Virus			15. NUMBER OF PAGES	
			16. PRICE CODE	
17. SECURITY CLASSIFICATION OF REPORT Unclassified	18. SECURITY CLASSIFICATION OF THIS PAGE Unclassified	19. SECURITY CLASSIFICATION OF ABSTRACT Unclassified	20. LIMITATION OF ABSTRACT	

Final Report for Phase III  
Contract No.: DAMD 17-91-C-1006  
Principal Investigator: Margo A. Brinton  
Date: July 31, 1993

Phase III of this contract required that we twice-clone selected simian hemorrhagic fever (SHF) virus-specific hybridoma cultures, expand two clones from each of the selected hybridomas and deliver to USAMRIID 10 vials of frozen cells of each clone as well as 50 ml of supernatant fluid from cultures of each clone. We have completed Phase III as described in detail below.

Production and screening of a second set of hybridomas. The set of SHFV hybridomas described in the Phase I and II Final Reports unfortunately did not continue to produce SHFV-specific antibody after single cell cloning. Therefore, a second set of hybridomas was produced. This time two sets of five mice were each immunized with SHF virions purified by centrifugation through a 15 to 55% sucrose density gradient. The first group of Balb c mice were 3 month old females and each received  $3.85 \times 10^7$  PFU of SHFV in 100  $\mu$ l by the subcutaneous route. The second group of Balb c mice were 1 month old females and each received  $9.5 \times 10^7$  PFU of SHFV in 100  $\mu$ l by the subcutaneous route. TiterMax (CytRx Corp.) was used as the adjuvant for the first virus injection. A second injection was given on day 32 to the first group of mice and on day 28 to the second group. For this injection mice in the first group were given pelleted virus resuspended in HBSS ( $9.5 \times 10^7$  PFU/100  $\mu$ l per mouse). Mice in the second group were given a 1:1 mixture of sucrose gradient purified virus and pelleted virus (a total of  $7.2 \times 10^8$  PFU/100  $\mu$ l per mouse). Immunized mice were bled approximately 21 days after the second injection and the plasma was tested for SHFV reactivity by ELISA assay at USAMRIID and by Western blotting at Georgia State University. Reactivity to SHFV Vp1, Vp2, and Vp3 proteins was detected in all 10 mice by Western blotting. Specific reactivity to purified virion as well as cell extract SHFV antigen preparations was observed in all mice by ELISA assay. The mouse in each group which showed the best response in both assays was selected as the spleen donor for the production of hybridomas.

Supernatants from the 268 wells in which cells grew were first tested for SHFV reactivity by ELISA. Those supernatants showing the highest levels of reactivity were subsequently tested by Western blotting. The results indicated that 6 of the supernatants reacted with Vp1, 6 of the supernatants reacted with Vp2, 7 of the supernatants reacted with a 35 KD and a 45 KD band in the Vp3 region and 5 of the supernatants reacted with a 35 KD band in the Vp3 region. At least three hybridomas representing each of the four unique reactivity patterns were selected.

Cloning of selected SHF virus specific hybridoma cultures. Western blot reactivity patterns are shown in Figure 1 and ELISA titers are shown in Table 1. The four selected hybridomas, AA4 (Vp2), AD4 (Vp3?), FC2 (Vp1), and HD4 (Vp3) (one for each reactivity pattern), were then subjected to two rounds of single cell cloning to assure the monospecificity and stability of the selected hybridoma cultures. Supernatants from cell positive wells in the single cell cloning dishes were first tested for SHFV reactivity by

ELISA. Approximately 90% of these wells produced antibodies with SHFV-specific reactivity. Those supernatants with the highest ELISA titers were selected (Table 2). In all cases the supernatants from these cloned cells reacted with the same SHFV protein as did the original hybridoma culture from which it was cloned. For each of the four unique reactivity patterns, two clones with good reactivity were selected and subjected to a second round of single cell cloning. Two clones of each reactivity type were again selected using the same methods as described above (Table 3).

Expansion of the selected, cloned hybridomas. The eight clones representing two clones for each of the four reactivity patterns were then expanded. Cells were aliquoted at a concentration of about  $10^7$  cells per vial and frozen by programmed reduction of temperature. Fifty milliliters of long-term culture fluid for each clone were also obtained. Ten vials of each type of cloned cell (80 vials in total) were shipped on dry ice and the supernatants were shipped on wet ice to Dr. Fred Knauert at USAMRIID.

Additional SHFV hybridomas. Although the shipment of hybridomas and supernatants already received by USAMRIID completely fulfills the contract, we have begun to twice-clone an additional backup hybridoma for each reactivity pattern. The backup set of four hybridomas selected for cloning are BB2 (Vp2), AC1 (Vp1), BB3 (Vp3?), and HC5 (Vp3). These hybridomas will be cloned twice and clones will be selected as described above. Eighty vials of frozen cells and 50 ml of supernatant from each of the 8 clones will then be sent to USAMRIID at no additional cost.

Accession For	
NTIS - CRA21	
DTIC - T-1	
U.S. Army - 100	
J. H. H. H. H.	
By	
Distribution	
Availability	
Dist	Availability Special
B-3	

RECEIVED 10/15/77

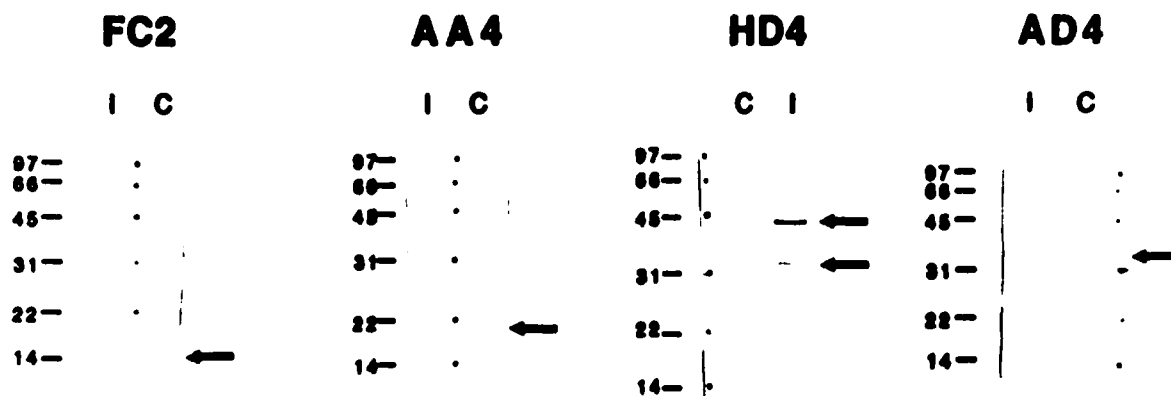


Figure 1. Western blot analysis of four selected SHFV hybridomas. FC2 reacts with Vp1. AA4 reacts with Vp2. HD4 reacts with a 45 KD and a 35 KD band. AD4 reacts with a 35 KD band. Arrows indicate positions of the virus protein bands. The positions of the molecular weight standards are indicated on the left side of each nitrocellulose strip. I - SHFV pelleted from infected tissue culture supernatant. C - uninfected pelleted tissue culture supernatant.

**Table 1.** The ELISA O.D. values, Western Blot and IgG Dot Blot results from the selected hybridoma supernatants.

Hybridoma	RV204	RV204	titer	SHF-HBSS	SHF-CIRL	WB protein	WB intensity	IgG intensity
AA4	0.60	50	0.00	0.00	VP2	unknown	strong	2
AD4	0.51	50	0.00	0.00	VP1	unknown	strong	4
FC2	0.55	50	0.00	0.00	VP3	unknown	strong	3
HD4	0.49	50	0.00	0.00	VP3	unknown	strong	4

**Table 2.** The ELISA O.D. values from once cloned hybridomas chosen to be cloned a second time.

A.	1st cloning	RV204 1:200				RV193 1:500				Pellet 1000			
		HD4/AB11	AD4/AA9	AA4/BB5	FC2/AA1	HD4/AB11	AD4/AA9	AA4/BB5	FC2/AA1	HD4/AB11	AD4/AA9	AA4/BB5	FC2/AA1
		1.68	1.30	1.40	1.18	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
B.	1st cloning	Dilutions of Supernatants				Diln.				Sum			
		5	20	80	320	5	20	80	320	5	20	80	320
	HD4/AB11	0.59 <sup>1</sup>	0.71	0.65	0.48	0.59 <sup>1</sup>	0.71	0.65	0.48	2.46	0.71	0.57	0.55
	AD4/AA9	0.57	0.62	0.45	0.31	0.57	0.62	0.45	0.31	1.94	0.57	0.55	0.55
	AA4/BB5	0.55	0.30	0.36	0.38	0.55	0.30	0.36	0.38	1.60	0.55	0.55	0.55
	FC2/AA1	0.24	0.48	0.17	0.16	0.24	0.48	0.17	0.16	1.50	0.40	0.40	0.40

<sup>1</sup> SHF gradient purified antigen was used at a dilution of 1:200.

**Table 3.** The ELISA O.D. values from the selected hybridomas after 2nd single cell cloning.

<u>2nd cloning</u>	<u>Dilutions of Supernatants</u>				<u>Iiter</u>	<u>Sum</u>	<u>O.D.</u>	<u>Diln.</u>
	<u>5</u>	<u>20</u>	<u>80</u>	<u>320</u>				
AA4/BB5-AC4	0.45 <sup>1</sup>	0.37	0.34	0.33	320	1.50	0.45	5
AA4/BB5-AE3	0.48	0.29	0.31	0.24	320	1.30	0.48	5
AD4/AA9-BE3	0.41	0.38	0.30	0.14	80	1.23	0.41	5
AD4/AA9-BF4	0.50	0.44	0.37	0.19	320	1.50	0.50	5
FC2/AA1-AF4	0.44	0.41	0.32	0.23	320	1.40	0.44	5
FC2/AA1-BH2	0.50	0.41	0.28	0.23	320	1.42	0.50	5
HD4/AB11-AA7	0.57	0.45	0.35	0.24	320	1.61	0.57	5
HD4/AB11-AD8	0.51	0.49	0.40	0.31	320	1.72	0.50	5

<sup>1</sup> SHF gradient purified antigen was used at a dilution of 1:200.